

Microinjection Method for *Anopheles gambiae* Embryos

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Materials

Name	Company	Catalog Number	Comments
10x Microinjection Buffer	-	-	1 mM NaHPO ₄ buffer, pH 6.8, 50 mM KCl
Blotting membrane (Zeta-Probe GT Genomic Tested Blotting Membrane)	Bio-Rad		Neatly and straightly cut into 2x1 cm piece
Conical tubes 50 ml (disposable centrifuge tube, polypropylene)	Fisher Brand		Ends cut
De-ionized or double-distilled water (ddH ₂ O)	Mili-Q		In a wash bottle
Dissecting microscope	Leica	Leica MZ12	For embryo alignment
Forceps			No. 5 size
Glass container	Pyrex	No. 3140	125 x 65
Glass slide	Fisher Brand	No. 12-549-3	75x26 mm
Incubator	Barnsted Lab-line	Model No. 150	28 °C
KCl			50 mM
Latex dental film	Crosstex International	No. 19302	
Microinjector	Sutter Instrument		XenoWorks Digital Microinjector
Microloader Pipette tips	Eppendorf		20 µL microloader epT.I.P.S.
Micromanipulator	Sutter Instrument		XenoWorks Micromanipulator
Micropipette	Rainin		20 µL
Micropipette puller	Sutter Instrument	Sutter P-2000 micropipette puller	
Microscope	Leica	DM 1000 LED or M165 FC	For microinjection
Minimum fiber filter paper	Fisher Brand	No. 05-714-4	Chromatography Paper, Thick
Mosquitoes	MR4, BEI Resources		<i>Anopheles gambiae</i> , mated adult females, blood-fed 4-5 days post-eclosion
NaHPO ₄ buffer			1 mM, ph 6.8
Nylon mesh			
Paint brush	Blick	No. 05831-7040	Fine, size 4/0
Petri dish			Plastic, (60x15 mm, 90x15 mm)
Sodium acetate			3M

Quartz glass capillaries	Sutter Instrument	No. QF100-70-10	With filament, 1 mm OD, ID 0.7 10 cm length
Water PCR grade	Roche	No. 03315843001	