

Video Article

Assays for the Specific Growth Rate and Cell-binding Ability of Rotavirus

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Keywords: Immunology and Infection, Issue 143, rotavirus, MA104 cell line, specific growth rate, cell-binding ability, plaque assay, RT-qPCR

Date Published: 1/28/2019

Citation: Kadoya, S.s., Sano, D. Assays for the Specific Growth Rate and Cell-binding Ability of Rotavirus. *J. Vis. Exp.* (143), e58821, doi:10.3791/58821 (2019).

Abstract

Rotavirus is the main etiological factor for infantile diarrhea. It is a double-stranded (ds) RNA virus and forms a genetically diverse population, known as quasispecies, owing to their high mutation rate. Here, we describe how to measure the specific growth rate and the cell-binding ability of rotavirus as its phenotypes. Rotavirus is treated with trypsin to recognize the cell receptor and then inoculated into MA104 cell culture. The supernatant, including viral progenies, is collected intermittently. The plaque assay is used to confirm the virus titer (plaque-forming unit: pfu) of each collected supernatant. The specific growth rate is estimated by fitting time-course data of pfu/mL to the modified Gompertz model. In the assay of cell-binding, MA104 cells in a 24-well plate are infected with rotavirus and incubated for 90 min at 4 °C for rotavirus adsorption to cell receptors. A low temperature restrains rotavirus from invading the host cell. After washing to remove unbound virions, RNA is extracted from virions attached to cell receptors followed by cDNA synthesis and reverse-transcription quantitative PCR (RT-qPCR). These protocols can be applied for investigating the phenotypic differences among viral strains.

Video Link

The video component of this article can be found at <https://www.jove.com/video/58821/>

Introduction

RNA viruses form a genetically diverse population, known as quasispecies¹, because of their mutation rate,² which is higher than that of DNA-based organisms. Population structure in quasispecies is affected by the population genetic factors, including mutation, selection pressure, and genetic drift. Strains within a single genetic lineage may show different phenotypes because of the genetic diversity. For example, Rachmadi et al. showed that free chlorine sensitivity was different among murine norovirus strains that originated from a plaque-purified strain S7-PP3³.

Rotaviruses (genus rotavirus in reoviridae family) are non-enveloped ds RNA viruses forming quasispecies². In addition to the population genetic factors described above, genome reassortment affects the genetic diversity of rotavirus because this virus has 11 segmented genomes⁴. Rotaviruses cause diarrhea mainly among infants, and infant deaths in 2013 were estimated about 250,000⁵. Two vaccines are in use in several countries and have been effective in reducing the burden of rotavirus infection, but some researchers are now discussing the presence of vaccine-escape mutants^{6,7,8,9}. The characterization of these mutants is important to understand the vaccine-escape mechanisms.

Here, we present protocols for two assays for evaluating the specific growth rate and cell-binding ability of rotavirus in order to understand the phenotypic differences among strains/mutants. The growth curve of rotaviruses has been presented in previous reports¹⁰, but growth parameters such as specific growth rate are not usually measured. A cell-binding assay conducted previously involves the immunofluorescent staining technique¹¹. We show here easier methods of using the plaque assay and RT-qPCR, which allow us to quantitatively discuss the difference in viral phenotypes. These methods are appropriate for the characterization of rotavirus phenotypes and may finally contribute to the construction of new vaccines effective for multiple genotypes.

Protocol

1. Medium Preparation

1. To make a cell culture medium (serum-containing medium), add 4.7 g of Eagle's MEM powder to 500 mL of distilled water. Autoclave at 120 °C for 20 min and let the medium cool to room temperature. Add fetal bovine serum (final concentration: 10%), L-Glutamine (2 mM), Penicillin Streptomycin (1%) and sodium bicarbonate (1.125 g/L). Store at 4 °C for 1 month.
2. Prepare the serum-free medium for the virus propagation as described in step 1.1, but without the fetal bovine serum. Store at 4 °C for 1 month.
3. For the plaque assay, sterilize 100 mL of Eagle's MEM medium (non-containing phenol red) by autoclaving. Let the medium cool to room temperature, and then add 2% FBS, 2% Penicillin Streptomycin, 4 mM L-Glutamine, and 2.25 g/L NaHCO₃. Store at 4 °C.

- For the plaque assay, sterilize 100 mL of 2.5% agarose gel by autoclave. Prepare the gel the same day the plaque assay is conducted. Store the gel at 47 °C in a water bath.

2. Cell Culture

- Remove a cryotube containing MA104 cell lines from the liquid nitrogen container. Place the cryotube in a water bath at 37 °C to thaw the cells. Add 1 mL of the cell suspension to 20 mL of the serum-containing medium in a T75 flask. Incubate the flask in an incubator at 37 °C and 5% CO₂ for 2 or 3 days.
NOTE: The final cell concentration in the suspension is about 10⁶ cells/mL.
- Once the cell monolayer reaches 80% confluency, remove the supernatant and wash the cells twice with 5 mL of 1x Dulbecco's PBS (phosphate buffered saline).
- Add 4 mL of 0.05% trypsin-EDTA to the flask and incubate at 37 °C for 5 min to detach the cells from the flask. Transfer the cell suspension to a 15 mL tube and centrifuge at 190 x g for 5 min.
- Discard the supernatant and resuspend the pelleted cells (10⁶ cells) in 1 mL of serum-containing medium prepared in 1.1. Dilute the resuspended cells at 100-fold with the medium.
- Add 3 mL of the diluted cell suspension to each well of 6-well (for the plaque assay) or 24-well plates (for the cell-binding assay), respectively. Incubate the plates in an incubator at 37 °C and 5% CO₂ under the saturated vapor for 2 or 3 days.
NOTE: A T75 flask is suitable to collect the time-course samples because the sample volume of the supernatant (1 mL) can be ignored compared to the total supernatant volume (30 mL). Meanwhile, the infectious titer of the virus in each supernatant is measured by the plaque assay, which is usually conducted using a 6-well plate. A 24-well plate is utilized for the cell-binding assay.

3. Specific Growth Rate of Rotavirus

NOTE: Rhesus rotavirus (RRV, genotype: G3P[3]) is utilized in this protocol because RRV can rapidly and easily form plaques with MA104 cells.

- Place a tube containing 1 mL of the virus suspension (10⁷ pfu/mL) in a serum-free medium stored at -80 °C in a water bath at 37 °C to thaw. Add 1 µg/µL trypsin from porcine pancreas to 1 mL of the virus suspension (final trypsin concentration is 4 µg/mL) and then vortex. Incubate the virus suspension at 37 °C and 5% CO₂ under the saturated vapor for 30 min.
NOTE: Trypsin from the other sources can be used, but the effect on rotavirus infectivity needs to be tested in advance.
- Dilute the activated virus suspension with a serum-free medium to adjust the multiplicity of infection (MOI) to 0.1 pfu/cell.
- Add 1 mL of diluted virus suspension to MA104 cell lines (80% confluent) in a T75 flask 3 days after the cell plating (2.1), incubate at 37 °C for 1 h, and gently shake the flask every 15 min.
- Then, add 30 mL of a serum-free medium containing 0.13 µg/mL of trypsin from a porcine pancreas to the flask. Incubate the flask at 37 °C and 5% CO₂ under the saturated vapor.
- Collect 1 mL of the supernatant in the flask at 0, 6, 12, 18, 24, and 36 (and/or 48) h post-infection (hpi) and replace the supernatant in the 1.5 mL tubes using a pipette.
- Conduct the freeze (-80 °C) and melt in a water bath at 37 °C cycle three times. Then centrifuge the tubes at 12,600 x g for 10 min at 4 °C. Collect the supernatant.
- Filter the supernatant with a distilled 0.2 µm filter to remove the cell fraction. Store the supernatant -80 °C in the refrigerator until applying it to the plaque assay for measuring the virus titer.
- Place the tubes containing the collected supernatant (step 3.5) in a water bath at 37 °C. Add 4 µg/mL trypsin to a 1 mL of 10-fold diluted sample and incubate at 37 °C for 30 min.
- During the 30 min incubation in 3.8, to begin the plaque assay for measuring the virus titer obtained from time course samples (step 3.5), wash the MA104 cells twice in a 6-well plate with 2 mL of 1x PBS after removing the serum-containing medium.
- Serially dilute the incubated samples with a serum-free medium and inoculate 1 mL of the diluted sample into each well. Incubate the plate for 90 min at 37 °C and 5% CO₂ under the saturated vapor, and gently shake the plate every 15 min.
- After incubation, remove the inoculum from the 6-well plate. Add 4 µg/mL trypsin to the medium prepared in (step 1.3). Gently but immediately add 3 mL of the medium mixed with agarose gel (the ratio is 1:1) to each well.
- Keep the plate at room temperature for more than 10 min (until the agarose gel becomes solid) and incubate for 2 days at 37 °C and 5% CO₂ under the saturated vapor.
NOTE: Pour the medium mixed with agar from the edge of the well.
- Add 1 mL of the 0.015% neutral red solution diluted with 1x PBS to each well and incubate at 37 °C and 5% CO₂ under the saturated vapor. Remove the dye after 3 h and incubate for 1 day at 37 °C and 5% CO₂ under the saturated vapor.
- The next day, count the number of plaques in each well and calculate the pfu/mL. Carefully check the cell confluence before the plaque assay to assure the plaque numbers.

4. Cell-binding Assay

NOTE: This protocol is based on Gilling's report¹³.

- Add 1 µg/µL trypsin from a porcine pancreas to 1 mL of the virus suspension (final trypsin concentration is 4 µg/mL) and then vortex (in the same manner as in 2.1). Dilute the virus suspension with a serum-free medium to adjust the MOI of 1 pfu/cell.
- Then, wash the MA104 cells twice on 24-well plate with 1 ml of Tris-buffered saline (TBS; 2.53 g/L Tris base, 6.54 g/L NaCl, 0.3 g/L KCl, 0.046 g/L Na₂HPO₄ to reach 1 L with distilled water).
- Inoculate 100 µL of diluted virus suspension to each well of a 24-well plate with cells, and incubate at 4 °C for 90 min, with gentle shaking every 15 min.

4. Remove the virus inoculum and wash the cells twice with 1 mL of TBS. To extract the double-stranded (ds) RNA of the rotavirus, add 140 μ L of 1x PBS and 560 μ L of the RNA extraction buffer (see **Table of Materials**) to each well. Mix adequately with a pipette (about 10x or until the haze or contaminant of cells in the buffer is not seen).
5. After recovering the double stranded RNA (dsRNA) according to the manufacturer's protocol, place the 1.5 mL tubes containing the dsRNA extract on a heat block at 95 $^{\circ}$ C for 5 min to denature the dsRNA, and then immediately place the tubes on ice and incubate for more than 2 min.
6. Synthesize the cDNA by using a reverse transcription kit (see **Table of Materials**). Add 4 μ L of denatured viral RNA solution to a PCR tube containing 16 μ L of mixture (**Table 1**) and mix it carefully with a pipette so as not to generate bubbles. Spin down the tubes.
7. Perform the reverse transcription with a thermal cycler under the condition shown in **Table 2**. If the cDNA is not used immediately, store the PCR tube containing the cDNA at -20 $^{\circ}$ C for up to 1 year.
8. Use the primers for quantitative PCR (Forward; 5'-ACCATCTACACATGACCCTC-3', Reverse; 5'-GGTCACATAACGCCCC-3')¹⁴ and a probe inserting a quencher (qPCR Probes; 5'-FAM/ATGAGCACA/quencher/ATAGTTAAAAGCTAACACTGTCAA/TAMRA/-3'), targeting the 963 – 1049 region of NSP3 genome segment of rotavirus (ST3 strain, GenBank: X81436).
NOTE: Quencher is inserted into the probe designed by Zeng et al.¹⁴
9. Dilute the standard plasmid serially (10^1 to 10^6 copies/mL) with PCR grade water to the qPCR mixture (**Table 3**) and make the master mix (20 μ L/sample) for qPCR following **Table 4**.
10. Add 20 μ L of the master mix to the well of a 96-well PCR plate, and then mix 5 μ L of cDNA samples or 5 μ L of the standard plasmid by pipetting 10 times. Start the reaction of the qPCR system according to the conditions shown in **Table 4**.
11. To calculate the rotavirus genome bound to the MA104 cell surface, conduct linear regression between Ct values and the known genome number of a standard plasmid, and estimate the sample's genome number. Then calculate the ratio of virion numbers binding to cells (G_t) to those in the initial inoculum (G_0).

Representative Results

An overview of two protocols for the specific growth rate and cell-binding assay of plaque- purified RRV strains is shown in **Figure 1A** and **2A**, respectively.

In the assay for the specific growth rate, the final virus titer reaches more than 10^7 pfu/mL when propagating on the T75 flask. If the maximum concentration is lower than 10^7 pfu/mL, the MA104 cell may not have become confluent or RRV was not activated well by trypsin. Some growth models are available for estimating the specific growth rate using the infectious unit data. In this protocol, the modified Gompertz model¹² is employed as an example;

$$\log \frac{N_t}{N_0} = A \cdot \exp \left\{ - \exp \left[\frac{\mu e}{A} (\lambda - t) + 1 \right] \right\}$$

where N_0 (10^4 pfu/mL in this study) and N_t (10^4 to 10^8 pfu/mL) are the virus infectious titer (pfu/mL) at 0 and t (example: 0, 6, 12, 18, 24, 36) hpi, respectively. A is the asymptotic value [$\log(N_{\infty}/N_0)$] (example: 3 to 4), μ is the specific growth rate [1/h], e is the Napier's constant and λ is the lag period [h]. Model parameters are obtained by the solver function of the analysis software, which minimizes the sum of squares of the difference between the observed and modeled values. In the example in **Figure 1B**, the specific growth rate (μ) is estimated to be 0.197 [1/h] and the lag period (λ) is 6.61 [h] by applying the least square method to a modified Gompertz model, and the relative virus titer at the stationary phase to the initial titer (log scale) (A) is 3.15 [$\log(N_{\infty}/N_0)$]. We have tested 6 rotavirus clones in total, and the estimated values of the specific growth rate ranged from 0.19 to 0.27 [1/h]. These estimated values are reliable because the coefficient of determination values in the model fitting is more than 0.98.

RRV virions binding to cell surfaces were about 10^3 copies/mL (binding efficiency was around 1%) when using a 24-well plate for the cell-binding assay (**Figure 2B**). The assay is usually conducted three times for every sample, and if a large variance in the copy number is observed in a sample, some problems such as over-washing and insufficient activation of RRV by trypsin may occur. The Ct value of qPCR exceeding about 36.0 is not preferable and is regarded to be below a detection limit in our qPCR condition.

	Volume/ 1 reaction
5 x PrimeScript Buffer	4.0 μ L
PrimeScript RT Enzyme Mix I	1.0 μ L
Oligo dT Primer	1.0 μ L
Random 6 mers	4.0 μ L
Deionized distilled water	6.0 μ L
ssRNA sample	4.0 μ L
Total	20.0 μ L

Table 1: Master mix composition for cDNA synthesis of rotavirus genome.

Temperature [°C]	Time
37	15 min
42	15 min
85	5 s
4	∞

Table 2: Reaction condition for cDNA synthesis of rotavirus genome.

	Volume/1 reaction
Premix Taq	12.5 µL
Forward primer (10 µM)	0.5 µL
Reverse primer (10 µM)	0.5 µL
Probe (10 µM)	0.5 µL
Reference Dye II	0.5 µL
Deionized distilled water	5.5 µL
cDNA sample	5.0 µL
Total	25 µL

Table 3: Master mix composition for quantitative PCR of rotavirus A genome.

Temperature [°C]	Time	
95	5 min	45 cycle
94	20 s	
60	1 min	
72	5 min	

Table 4: Reaction condition for quantitative PCR of rotavirus A genome.

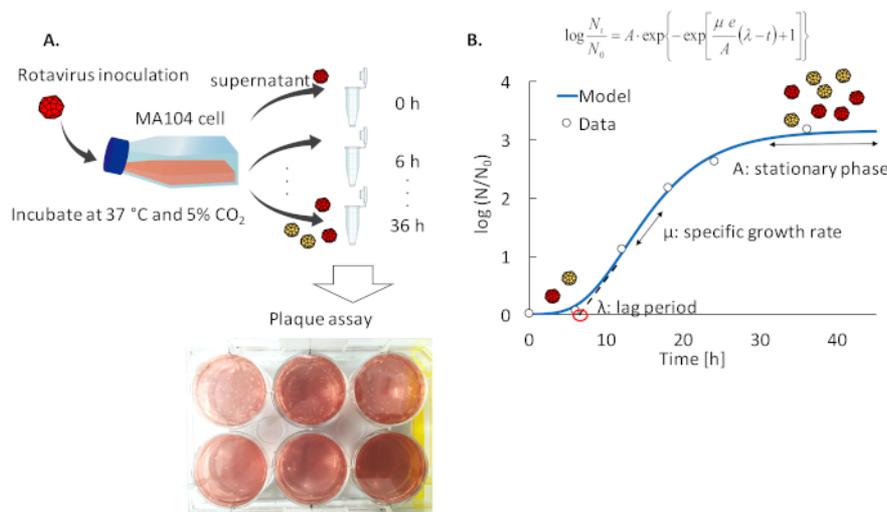


Figure 1: Schematic overview of the estimation of rotavirus growth and the growth curve of rotavirus. (A) The infectious unit of rotavirus is measured with the plaque assay. (B) The curve (blue line) was approximated by the modified Gompertz model based on observed data in our laboratory (white circle). The specific growth rate [μ]; 0.197 [h^{-1}], lag period (λ); 6.61 [h], the relative virus titer at the stationary phase to the initial titer (log scale) (A); 3.15 [$\log(N_{\infty}/N_0)$]. [Please click here to view a larger version of this figure.](#)

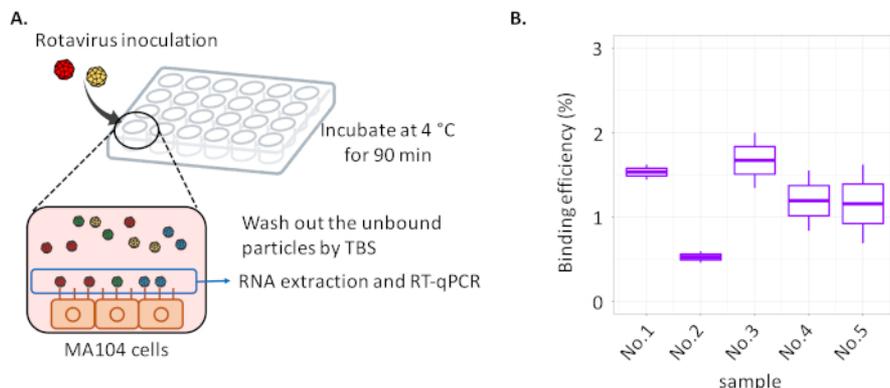


Figure 2: Schematic overview and representative result of the cell-binding assay of five RRV strains purified from plaques in our laboratory. (A) A cell culture plate inoculated with rotavirus is incubated at 4 °C for inhibiting the virus invasion into cells. After incubation and removing the unbound viral particles to cells, quantify the number of genomes originating from bound viral particles to the cell surface with RT-qPCR. (B) The result of the cell-binding assay was displayed as binding efficiency (%), which was the ratio of bound viral particles to those present in the inoculum. Bold bar: median, end of boxes: quartile deviation, end of line: maximum and minimum. [Please click here to view a larger version of this figure.](#)

Discussion

Our protocol for measuring the specific growth rate is easier than previous ones and can be adapted for other viruses unless their cell culture system has not yet been established. In this study, we used RRV (G3P[3]) because this strain can form plaques easier than human rotaviruses when using MA104 cell lines. Some human rotavirus strains cannot form plaque in this cell line. Therefore, instead of the plaque assay, the focus forming unit (FFU) assay¹⁵ or median tissue culture infectious dose (TCID50) assay can be applied for many rotavirus strains¹⁶. The presented protocol for determining the specific growth rate can be used for other virus types but is not suitable for viruses for which no established cell culture system is established. Before starting the experiment for the specific growth rate, it is better to know in advance when the virus infectious titer starts to increase and reaches the stationary phase in a preliminary test. If too many plaques are present, the plaque assay should be conducted again after changing the dilution rate of the virus samples. The hours post-infection (hpi) to collect samples are also important because the slope of the exponential growth phase may be underestimated if the proper time point to reach the stationary phase is missed. In the approximation by the modified Gompertz model, a coefficient of determination should always be calculated and checked. If the fitness to the modified Gompertz model is low, other growth models such as the modified logistic model¹² may be preferable.

In handling cells, when removing the medium or virus inoculum, the PBS for washing or agarose gel for plaque assay must be promptly added to each well of a cell culture plate to prevent the cells from dryness. At the same time, you must gently pour PBS or agarose to cells not to detach from wells. This step is the most important in both assays (step 3.9 and 3.11). If the plaque assay has too many samples, we recommend subdividing the agarose gel (100 mL each maximum is recommended) into several medium bottles and keeping the bottles warm until just before use since agarose gel is solidified within about 10 min at room temperature. Since trypsin is vulnerable to high temperatures¹⁷, a trypsin solution should be added to a medium and agarose for the plaque assay after adequately cooling down.

In the cell-binding assay, the incubation for virus binding to cells must be done at 4 °C to prevent invasion of the cells. According to Gilling's method¹³, cells are prone to drying at low temperatures, so gentle shaking is necessary every 15 min during incubation. The RNA extraction kit utilized here can be substituted for other kits. The slope of the standard curve in RT-qPCR should be approximately 3.3, and the coefficient of determination should be more than 0.98. Compared to the fluorescence microscope to visualize the localization of viruses in cells, the assay is more rapid and easier to use because binding of fluorescent substances to viruses is not necessary.

Recently, human intestinal enteroid (HIE), exhibiting a similar cellular composition and function as human gastrointestinal epithelium, has become available for rotavirus propagation¹⁸. The use of HIE may enable us to evaluate the specific growth rate and cell-binding ability of non-culturable strains of rotavirus. Also, both experiments described here may be applied to the evaluation of drug effects on both phenotypes¹⁹. The protocols presented here make it possible to quantitatively discuss the changes in phenotype parameter values of rotavirus strains under varied conditions.

Disclosures

The authors have nothing to disclose.

Acknowledgments

This work was supported by "The Sanitation Value Chain: Designing Sanitation Systems as Eco-Community Value System" Project, ResearchInstitute for Humanity and Nature (RIHN, Project No.14200107).

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