

Materials List for:

# Visualizing Bacteria in Nematodes using Fluorescent Microscopy

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## Materials

Name	Company	Catalog Number	Comments
Lipid Agar (sterile)			8 grams nutrient broth, 15 grams agar, 5 grams yeast extract, 890 ml water, 10 ml 0.2 g/ml MgCl <sub>2</sub> · 6H <sub>2</sub> O, 96 ml corn syrup solution*, 4 ml corn oil* Stir media while pouring plates *add sterile ingredient after autoclaving
Corn Syrup Solution (sterile)			7 ml corn syrup, 89 ml water mix and autoclave
Egg Solution			16.6 ml 12% sodium hypochlorite, 5 ml 5M KOH, 80 ml water
Lysogeny Broth (sterile)			5 grams yeast extract, 10 grams tryptone, 5 grams salt, 1 L water mix and autoclave
Microfuge	Fisher	13-100-675	Any microfuge that holds microfuge tubes will work
Centrifuge	Beckman	366802	Large table top centrifuge that holds 15 ml and 50 ml conical tubes
Sterile 60 mm X 15 mm Petri Dish	Fisher	0875713	
50 ml centrifuge tubes	Fisher	05-539-6	
15 ml centrifuge tubes	Fisher	05-531-6	
Sterile 100 mm X 20 mm Petri Dish	Fisher	0875711Z	Deeper than standard Petri dishes
24-well plate	Greiner Bio-One	662000-06	
Microscope			The microscope needs florescent capabilities compatible with your fluorophore
Paraformaldehyde	Electron Microscopy Sciences	15710	
PBS (sterile)			8 g NaCL 0.2 g KCL 1.44 g Na <sub>2</sub> HPO <sub>4</sub> 0.24 g KH <sub>2</sub> PO <sub>4</sub> 1 L water Adjust to a pH of 7.4 and water to 1 L and autoclave
Microfuge tubes	Fisher	05-408-138	2 ml or 1.5 ml tubes
Shaker			Any shaker that causes the liquid to gently move will work
Diaminopimelic acid	Sigma	D-1377	If needed, supplement media to a concentration of 1 mM